

Is all seagrass habitat equal? Seasonal, spatial, and interspecific variation in productivity dynamics within Mediterranean seagrass habitat

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Abstract

Seagrass meadows' ability to capture carbon through actively sequestering autochthonous carbon via photosynthesis means they represent a nature-based solution to rising carbon emissions. In multispecies seagrass communities, and due to species introduction or predicted range shifts, it is important to know which species deliver different carbon sequestration gains to inform conservation actions. Large benthic chamber (volume = 288L) experiments assessed the seasonal and spatial metabolism dynamics of the endemic and dominant Mediterranean seagrass *P. oceanica*, whilst small benthic chamber (volume = 7L) experiments compared the dynamics between *P. oceanica*, the native *C. nodosa* and non-native *H. stipulacea*. Within shallow *P. oceanica* edge habitat lower Net Apparent Productivity (NAP) occurs in autumn ($\bar{x} = 1.4$, $SD \pm 3.24$ O₂ mmol m⁻² d⁻¹) compared to summer ($\bar{x} = 10.9$, $SD \pm 3.03$ O₂ mmol m⁻² d⁻¹) corresponding with periods of light limiting and light saturating conditions, but it remains overall autotrophic annually (2.6 C mol m⁻² yr⁻¹). However, spatial heterogeneity exists, core areas of *P. oceanica* were more productive ($\bar{x} = 21.7$, $SD \pm 4.21$ O₂ mmol m⁻² d⁻¹) compared to edge habitat with spatial changes in productivity relating to plant surface area (96%), shoot density (81%), blade length (72%) and seagrass percentage cover (64%). In fact, patchy short *P. oceanica* has similar NAP ($\bar{x} = -3.6$, $SD \pm 8.9$ O₂ mmol m⁻² d⁻¹) to the non-native *H. stipulacea* ($\bar{x} = 2.1$, $SD \pm 4.7$ O₂ mmol m⁻² d⁻¹). Yet *H. stipulacea* has a significant metabolic advantage over *C. nodosa* (NAP $\bar{x} = 0.0004$, $SD \pm 0.0011$ O₂ mmol cm⁻² d⁻¹ vs $\bar{x} = -0.0012$, $SD \pm 0.0007$ O₂ mmol cm⁻² d⁻¹). This study demonstrates that not all seagrass habitat is equal. If seagrass meadows are to play a part in mitigating CO₂ emissions, variability in primary productivity within seagrass meadows needs to be accounted for to produce comprehensive autochthonous carbon sequestration estimates. This means seagrass meadow species composition, the condition of these species and the potential interactions between seagrass species must be better understood.

1 Introduction

Seagrass meadows act as a major global carbon sink (Duarte et al., 2013), therefore restoration or expansion of seagrass beds represent a potential nature-based solution to rising carbon emissions. Seagrasses capture carbon through actively sequestering autochthonous carbon by photosynthesis and passively trapping allochthonous carbon within their architectural structure. Allochthonous carbon is typically considered more labile, therefore deposits of autochthonous carbon are those expected to lead to long-term stable carbon deposits (Mazarrasae et al., 2018). The metabolic rates of global seagrass communities favour net autotrophy, with temperate meadows typically favoured to have a higher net autotrophy than tropical meadows (Duarte et al., 2010), suggesting not all seagrass is equal in its ability to sequester carbon.

The temperate seagrass *Posidonia oceanica* forms vast monospecific meadows in the Mediterranean and is unique in its ability to form vertical mattes that can store sedimentary carbon for millennia (Mateo et al., 1997). It is perhaps why some herald these meadows to represent the global maximum in carbon sequestration among seagrasses (Lavery et al., 2013). Unlike terrestrial soils, *P. oceanica* sediments do not become saturated with carbon over time because they can accrete vertically. If the vertical accretion matches the rate of sea level rise, they potentially have a limitless capacity, which in part demonstrates their suitability for climate mitigation policy efforts (Howard et al., 2017). However, *P. oceanica* meadows have undergone severe regression in the last 50 years (34% loss), with only localised areas of persistence and growth (Telesca et al., 2015). The causes of decline for *P. oceanica* include water quality degradation, coastal modification, mechanical damage (i.e., bottom trawling, anchoring, mooring, culture farm occupation), extreme weather events and non-native macroalgae invasion (Santos et al., 2019).

At the Mediterranean scale, *P. oceanica* populations in the Western and Eastern basins maintain genetic differentiation due to present-day dispersal limits (Arnaud-Haond et al., 2007). There is a greater distribution of *P. oceanica* in the Eastern basin (713,992 ha) compared to the Western basin (510,715 ha) (Telesca et al., 2015). Despite the genetic variation between basins and larger coverage in the Eastern Mediterranean basin, the majority of *P. oceanica* metabolism estimates have come from the Western Mediterranean basin (Frankignoulle and Bouqueneau, 1987; Holmer et al., 2004; Gazeau et al., 2005; Barron and Duarte, 2009; Olive et al., 2015; Champenois and Borges, 2018). There is a single published study on *P. oceanica* metabolism in the Eastern Mediterranean basin, in the western region of the Aegean Sea (Apostolaki et al., 2010). Metabolism and carbon sequestration estimations for *P. oceanica* are considered one of the most well researched amongst seagrass species (Nordlund et al., 2018), yet there are distinct local knowledge gaps and spatial biases, particularly within the Eastern Mediterranean basin that need to be addressed. In the Eastern Mediterranean basin, the sea surface has warmed by 0.05 ± 0.009 °C yr⁻¹ compared to just 0.03 ± 0.008 °C yr⁻¹ in the Western Mediterranean basin from 1985 to 2006 (Nykjaer, 2009). Given the Eastern Mediterranean basin is warming faster than the Western Mediterranean basin understanding if the metabolism and drivers of productivity are different in Western Mediterranean seagrass is important.

Seagrass metabolism is influenced by multiple variables including light (Champenois and Borges, 2018), nutrient availability (Holmer et al., 2008; Apostolaki et al., 2010), temperature and ocean acidification (Berg et al., 2019); which leads to seasonal fluctuations in productivity. Seagrass depth distribution is determined by light availability, as under insufficient light conditions the plant does not meet the photosynthetic requirements needed to maintain positive metabolic and carbon balance (Ralph et al., 2007), moving away from a state of net autotrophy and net carbon storage. However, high light intensity also causes photodamage reducing the photosynthetic efficiency and causing

photoinhibition. Subsequently, maximum annual solar irradiance does not necessarily coincide with annual maximum *P. oceanica* gross primary productivity (GPP) (Champenois and Borges, 2018). The optimal thermal conditions for *P. oceanica* are between 17 - 20 °C (Champenois and Borges, 2018), therefore its metabolism both above and below these conditions, is not optimal for maximum oxygen production and is more likely to become heterotrophic. Annual patterns of *P. oceanica* metabolism therefore comprise periods that alternate from negative to positive carbon balance as temperature and light availability change with seasons (Alcoverro et al., 2001). If metabolism estimates were produced only during the summer months it would overestimate the meadow's net carbon sequestration capacity (Champenois and Borges, 2012). Notably as stable carbon stocks rely on autochthonous carbon deposits, we will also assess key environmental parameters that influence photosynthesis in these different seasons: change in light and temperature.

Seagrass meadows are not always a uniform habitat and whilst *P. oceanica* meadows can form continuous meadows, but patchy coverage across meadows is common. Patchy *P. oceanica* creates complex seascapes, which are mosaics including habitats of sand, *P. oceanica* dead matte and live *P. oceanica* (Borg et al., 2006; Abadie et al., 2015). This complexity also acts within a *P. oceanica* meadow, because at the junction between the seagrass and adjacent habitats there are considerable edge effects, resulting in distinct areas of central and edge *P. oceanica* habitat (Abadie et al., 2018). Meadows that experience a wave exposure gradient from low to high energy develop patchier dynamics (Folkard, 2005; Pace et al., 2016). Anthropogenic factors have also increased patchiness including anchoring, impact from historic military activity, fishing practices and fish farming (Montefalcone et al., 2009; Abadie et al., 2015). When a *P. oceanica* meadow is described as patchy it has lower overall cover, more complex patch shapes and reduced within-patch architectural complexity (Pace et al., 2016). This patchiness influences the available surface of photosynthetically active plant material. It is therefore important that in this study we consider the spatial variation in canopy architecture of *P. oceanica* when considering its carbon sequestration potential.

The Mediterranean temperate-tropical combination of seagrass species is considered a unique bioregion of seagrass diversity (Short et al., 2007). Alongside the dominant endemic *P. oceanica*, the next most prevalent species in the Aegean Sea is the native *Cymodocea nodosa*. The eastern region of the Aegean Sea sits at a crossroad for alien species expansion (Pancucci-Papadopoulou et al., 2012), including the non-native seagrass *Halophila stipulacea*, introduced to the Mediterranean and first reported in 1894 off the Island of Rhodes in the south-eastern region of the Aegean Sea (Fritsch, 1895; also see *Halophila decipiens*, Gerakaris et al., 2019). *H. stipulacea* was listed amongst the '100 Worst Invasive Species' in the Mediterranean (Streftaris and Zenetos, 2006). However, there are discrepancies in whether it should be considered 'invasive' or not as no ecological consequences of its introduction and spread in the Mediterranean have been reported (Williams, 2007). Scarce research has focused on its potential impact or contribution to ecosystem services within Mediterranean coastal ecosystems, yet *H. stipulacea* habitat was recently suggested to support carbon sequestration (Apostolaki et al., 2019; Wesselmann et al., 2021). Given *H. stipulacea* is one of the longest monitored non-native species in the Mediterranean, there has been a clear lag from reporting its presence and rate of expansion, to understanding its impact within the communities where it has successfully established.

In the Mediterranean *H. stipulacea* can be found in single species meadows, multi-species meadows with the native *Cymodocea nodosa*, in the free spaces between patches of *P. oceanica* or in habitats previously devoid of seagrass (Boudouresque et al., 2009). Whilst there are reports of its 'invasive' behaviour on sandy substrata when it exists in high abundances, no displacement of native species has yet been reported (Tsiamis et al., 2010). Given that *H. stipulacea* has various contexts in which it

can be found in the Mediterranean its contribution to community productivity may be context dependent. When *H. stipulacea* has colonised areas previously absent of seagrass its presence has increased the distribution of seagrass habitat in the Mediterranean, this is one reason why it can be considered a new potential blue carbon sink habitat (Apostolaki et al., 2019; Wessellmann et al., 2021). The potential of *H. stipulacea* to also influence the net productivity of mixed seagrass habitats, especially considering evidence it can colonise patchy areas of bare ground where *P. oceanica* may have been present in the past (Telesca 2015), will be assessed in this study.

We will determine seasonal metabolism measurements for the dominant seagrass *P. oceanica* in the eastern region of the Aegean Sea and the relative influence of seasonal changes in light, temperature, and seagrass canopy height, on net productivity. The accumulative seasonal estimates will then provide a conservative estimate of annual meadow carbon sequestration potential. Alongside this spatial variation in productivity between edge and central *P. oceanica* habitat will be assessed, together with the potential of various plant biometrics to describe spatial plant productivity. Finally, species-specific metabolism from both native seagrass species *P. oceanica* and *C. nodosa* will be placed in direct comparison to the non-native seagrass species *H. stipulacea*. Our results will contribute to suitable Mediterranean scale carbon budgets and to begin to assess potential changes that may take place in the future given current climate change predictions.

2 Methods

2.1 Study site

This study took place at Vroulia Bay (37.317460° N, 26.724704° E), Lipsi Island, which is part of the Dodecanese Islands, in the eastern region of the Greek Aegean Sea. This island suited the following study due the accessibility and distribution of the three seagrass species *P. oceanica*, *C. nodosa* and *H. stipulacea*. Vroulia Bay is sheltered with minimal human presence. The bay consists of a multispecies seagrass meadow, largely dominated by monospecific areas of *P. oceanica*. However, the bay also houses small monospecific patches of *C. nodosa* and *H. stipulacea*, as well as areas where two or all three of the species form mixed meadows.

2.2 Experimental Design

This metabolism study exists in two parts. Firstly, and primarily, large benthic chamber experiments focus on the seasonal and spatial metabolism dynamics of *P. oceanica*, these took place from autumn 2018 to summer 2019; seasonal *P. oceanica* sampling November 3rd – 9th, April 12th – 19th and July 2nd – 12th; spatial *P. oceanica* sampling July 2nd – August 12th. Secondly small benthic chamber experiments focus on comparative metabolism dynamics between *P. oceanica*, *C. nodosa* and *H. stipulacea*, collected 20th June – 17th July 2019.

2.2.1 Seasonal and spatial *P. oceanica* metabolism -large benthic chamber setup

The large dome shaped clear PVC benthic chambers (diameter = 1 m, height = 50 cm, benthic surface area = 0.79 m², volume = 288 L) were deployed by free divers (Fig. 1) in the shallow areas of Vroulia Bay 1.6 - 2 m depth amongst monospecific areas of *P. oceanica*. PME miniDOT® loggers were fitted to the central (highest) part of the dome chamber, to record dissolved oxygen and temperature in 10 minutes intervals. HOBO Pedant® loggers were placed within the chambers on the seafloor to record relative irradiance in 5 minutes intervals. Benthic chambers were fitted with a pump secured to the inside of the chamber roughly 30 cm from the seafloor. This pump circulated the water within the dome at a flow rate of roughly 480 L hr⁻¹ in 10 minute intervals (10 minutes on

170 followed by 10 minutes off), as such the overall flow rate within the chamber was considered to be
 171 240 L hr⁻¹. A thin cable went from the pump, within the benthic chamber, to an external submersible
 172 battery pack. Given the morphology and typically large aboveground biomass of *P. oceanica*, the
 173 large benthic chambers combined with internal mixing support sufficient volume of water inside the
 174 chamber relative to vegetative biomass to prevent oxygen saturation (Olive et al., 2015), and support
 175 accurate measurement of oxygen by the sensors (Annex 1). The benthic chambers were held in place
 176 by large chains and sank 5 cm into the sediment to create a seal. The benthic chambers were
 177 deployed within the same 2 hour timeframe each morning (10:45 am -12:45 pm) then left *in situ* for
 178 24 hour (autumn and summer) or 23 hour (spring) incubations.

179 The large *in situ* benthic chambers were deployed over edge habitat in autumn, spring and summer.
 180 Patchy edge habitat is considered to be the edge of large *P. oceanica* areas (> 2m) or the edge and
 181 centre of small *P. oceanica* patches (< 2m). Patches of *P. oceanica* < 2m in size are assumed to
 182 consist entirely of edge habitat due to the proximity of the centre to the edge. Edge habitat creates a
 183 good seal at the base of the large benthic chambers because of the high proportion of sand to low
 184 proportion of rhizome. This avoids damage to the rhizome and roots from the chamber.
 185 Simultaneously a large control benthic chamber was placed over the adjacent unvegetated sandy
 186 control habitat. If it was not possible to simultaneously deploy the control chamber, they were
 187 deployed at the next sampling opportunity the day after the first incubation is completed.

188 To achieve a comparison between more fragmented edge seagrass habitat and more continuous
 189 central meadow habitat, in summer domes were placed over *P. oceanica* within the meadow centre in
 190 addition to those deployed over edge habitat. Within dense areas of *P. oceanica* the rigid PVC
 191 benthic chamber likely cut the rhizomes and roots, however the stress caused is generally considered
 192 marginal (Champenois and Borges, 2012). Control chambers were not placed out simultaneously to
 193 the central meadow measurements, as control measurements from the edge habitat studies (e.g., bare
 194 sand, n = 4) were already available.

195 **2.2.2 Species specific productivity -small benthic chamber setup**

196 An area within Vroulia bay was chosen where all three species were in proximity, therefore
 197 maintaining as much as possible similar *in situ* environmental conditions for each species (appx. 7m
 198 depth). An area of patchy mixed meadow transitioned in areas from *P. oceanica* dominant to
 199 *C. nodosa* dominant, with scarce intermittent presence of clumps of *H. stipulacea*, as such this was
 200 chosen for the metabolism measurements focusing on *P. oceanica* and *C. nodosa*. Care was taken to
 201 avoid placing benthic chambers over mixed species compositions and generally the benthic chambers
 202 only encompassed the intended dominant species. Two of five *C. nodosa* chambers covered a small
 203 amount of either *P. oceanica* or *H. stipulacea*. Nevertheless, in those instances *C. nodosa* represented
 204 93% and 96% of the total seagrass plant surface area within the chamber and therefore these two
 205 replicates were still considered representative of *C. nodosa* patches. One of six *P. oceanica*
 206 chambers covered a small amount of *H. stipulacea* but *P. oceanica* represented 98% of the total plant
 207 surface area within the chamber and therefore was also considered representative of *P. oceanica*.
 208 Directly adjacent to the *P. oceanica* and *C. nodosa* was a monospecific area of *H. stipulacea*, which
 209 was utilised for the metabolism measurements focused on this species.

210 Three smaller clear dome PVC benthic chambers (diameter = 30 cm, height = 15 cm, benthic surface
 211 area = 0.071 m², volume = 7 L) were deployed by free divers, secured with a small chain and sank
 212 into the sediment to create a seal (Fig. 2). The benthic incubations took place for each seagrass
 213 species consecutively; *H. stipulacea* 20th June – 5th July (n = 6), *C. nodosa* 10th – 14th July (n = 5) and
 214 *P. oceanica* 14th – 17th July (n = 6). An unvegetated control chamber was simultaneously deployed

every day. The same oxygen, light and temperature loggers and temporal sampling methods were used as per the large domes described above. The benthic chambers were deployed within the same half an hour timeframe each morning (11:00 - 11:30 am) and left *in situ* for ~ 23 hours. As *H. stipulacea* and *C. nodosa* are smaller seagrass species with lower above ground biomass, the small benthic chamber design should sufficiently provide a suitable volume of water to vegetative biomass to prevent oxygen saturation. The *P. oceanica* within this area of the bay is naturally patchy and short, which should also accommodate the smaller chamber size and volume (Fig. 2B).

2.2.3 Quantifying seagrass canopy within the benthic chamber

The quantity of seagrass canopy enclosed within all benthic chambers was quantified; large benthic chambers a representative sample was selected, measured *in situ* and scaled up to reflect the full chamber; small chambers had all the leaf material collected and measured back at the lab.

For all large benthic chambers, several measurements were taken from the seagrass within the benthic chambers; an estimate of *P. oceanica* cover (%); shoot density (Sht_{dens}) (m^{-2}) within three 20 x 20 cm quadrats randomly placed on *P. oceanica*; the number of blades (B_{Number}) ($shoot^{-1}$) from 5 - 6 randomly selected shoots; canopy blade length (B_{length}) and blade width (B_{width}) (cm) from 5 – 6 randomly selected blades.

Plant surface area (m^2) for each chamber was calculated from the seagrass biometrics:

$$Shoot_{Surface\ Area} = 2(B_{length} \times B_{width}) \times B_{Number}$$

$$Plant\ surface\ Area = Shoot_{Surface\ Area} \times \left(Sht_{Dens} \times \frac{Cover}{100} \right)$$

For the small benthic chambers all leaf material from within the chambers was collected by cutting the leaves leaving the rhizome and roots, this along with the small sample size within these chambers' limits damage to the overall sampling area. The aboveground leaf material was transported back to the lab where they were separated from any dead leaf litter and all individual leaves were photographed using a Canon EOS 750D DSLR camera, on a white background. The planar area of the seagrass leaves within corresponding benthic chambers were determined, by Image J software, doubling to determine total leaf surface area (modified from, Schneider et al. 2012) and converted into areal based estimate ($cm^2\ m^{-2}$) using the chamber benthic surface area.

2.3 Community metabolism calculations

There is no need to consider diffusive exchange with the atmosphere as benthic incubations were carried out in closed systems. The record from the PME oxygen logger was used to calculate net community productivity (NCP) for every 10-minute interval over each 24-hour period (modified from Cole et al., 2000):

$$\Sigma \Delta DO = NCP_{Daily}$$

where DO is dissolved oxygen. This was applicable to the autumn and summer large benthic chamber *P. oceanica* samples when 24-hour incubations were completed.

HOBOPendant® light intensity data was used to distinguish the length of the night period. The total hours of daylight (H_d) were determined as:

$$H_d = 24 - H_n$$

where H_n is the total hours of night-time darkness.

The spring *P. oceanica* and mixed species incubations were completed across 23-hour incubations missing one hour of daylight incubation. $NCP_{Daytime}$ (Champenois and Borges, 2012; Rodriguez et al., 2016) was calculated whereby t is the daylight incubation time:

$$NCP_{Daytime} = \left(\frac{\Sigma \Delta DO}{t} \right) \times H_d$$

During darkness GPP is 0, therefore the change in dissolved oxygen in any 10-minute interval is assumed to be a result of community respiration. The change that occurred during the night-time dark period were summed to calculate CR_{Night} :

$$\Sigma \Delta DO = CR_{Night}$$

The combination of the net community change during the daytime ($NCP_{Daytime}$) and night-time dark period (CR_{Night}) determine NCP_{Daily} for 23-hour incubations made during Spring:

$$NCP_{Daily} = NCP_{Daytime} + CR_{Night}$$

Since the benthic chamber incubations do not acquire a direct measurement of CR_{Day} , we assume the hourly value of CR_{Night} and CR_{Day} are equal (Cole et al., 2000):

$$CR_{Day} = \left(\frac{CR_{Night}}{H_n} \right) \times H_d$$

This allows daily community respiration (CR_{Daily}) to be found:

$$CR_{Daily} = CR_{Day} + CR_{Night}$$

NCP_{Daily} is then calculated as:

$$NCP_{Daily} = GPP_{Daily} - CR_{Daily}$$

All metabolism measurements (O_2 mg L^{-1} d^{-1}) are converted into areal based estimates (Olive et al., 2015), whereby the volume (V) and area (A) is relative to the benthic chamber setup:

$$O_2 \text{ mmol } m^{-2} d^{-1} = \frac{\left(\left(\frac{DO}{32} \right) \times V \right)}{A}$$

The corresponding GPP:CR ratios were calculated:

$$\frac{GPP}{CR}$$

The photosynthetic and respiratory quotient of 1 mol of O_2 : 1 mol CO_2 is applied to terrestrial plants that use starch and sugars as respiratory substrates, as observed in *P. oceanica* (Alcoverro et al.,

2001). The photosynthetic and respiratory quotient of 1 mol of O₂: 1 mol CO₂ is conservatively applied to a wide number of seagrass species (Duarte et al., 2010) and therefore also used for *H. stipulacea* and *C. nodosa*. Therefore, the metabolism values are interchangeable as units of oxygen or carbon (O₂ mol m⁻² d⁻¹ and C mol m⁻² d⁻¹).

2.4 Net Apparent Productivity calculations

The net apparent productivity (NAP) of the seagrass and epiphytes (modified from Murray and Wetzel, 1987), was calculated as the difference between estimates for the NCP of the seagrass benthic chambers and the average NCP of the sediment and plankton in the unvegetated control chambers (Murray and Wetzel, 1987) from the same corresponding season. This calculation assumes the only difference between the vegetated and unvegetated benthic chambers is the presence of the seagrass and epiphytes.

The expected contribution of epiphyte productivity is deemed minimal due to the overall high biomass of *P. oceanica* to that of the epiphytes (Cox et al., 2015), therefore the large benthic chamber NAP primarily reflects the productivity of the *P. oceanica*. To account for seasonal changes in seagrass canopy the seasonal NAP measurements were also presented separately standardised by the canopy height with the large benthic chambers (O₂ mmol m⁻² cm⁻¹ d⁻¹).

To determine the NAP of the small benthic chambers the average NCP of the unvegetated control chambers deployed at the same time as the corresponding seagrass species were used. Given each seagrass species were deployed consecutively the corresponding unvegetated controls were grouped thus; 20th June – 5th July (n = 6) corresponding to *H. stipulacea*, 10th – 14th July (n = 3) for *C. nodosa* and 14th – 17th July (n = 3) for *P. oceanica*. These comparisons of species NAP were also presented separately standardised by the respective plant surface area (O₂ mmol cm⁻² d⁻¹), calculated with the scaled-up plant surface area from within the small benthic chambers (cm² m⁻²).

Accounting for the metabolism of the community within a seagrass meadow is important to determine the net productivity at a community level, but as the autotrophy within the community is largely attributed to the primary productivity of the seagrasses (Hemminga and Duarte, 2000), distinguishing the NAP of the *P. oceanica* enables a clearer determination of the influences driving this metabolic component of the community. Justifying the approach to distinguish between NCP (proxy of community metabolism) and NAP (proxy of plant productivity) within the *P. oceanica* meadow ecosystem, as seasonal community metabolism may partially mask the drivers in *P. oceanica* productivity. However, NCP estimates are widely accepted for benthic chamber metabolism studies within seagrass meadows (Duarte et al., 2010; Olive et al., 2015), because they clearly quantify the extent to which the overall community is a carbon sink or source. Hence, we also chose to present our results in NCP as it allows for closer comparisons to the existing *P. oceanica* NCP estimates.

2.5 Data analysis

2.5.1 Seasonal *P. oceanica* NCP and NAP

The three samples from winter and spring and four samples in summer, meant finding an appropriate data distribution to the seasonal *P. oceanica* benthic chamber data is challenging, but Shapiro-Wilk tests confirmed that when grouped by Season the NCP and NAP does conform to normal distribution (autumn $W = 0.9903741$, $P > 0.05$; spring $W = 0.8167365$, $P > 0.05$; summer $W = 0.9591216$, $P > 0.05$;) (ANNEX 2). Whilst Bartlett's test confirmed equal variances across the NCP and NAP data (K

= 0.00952, $P > 0.05$). Therefore, ANOVAs were applied to assess if there was a significant seasonal difference in *P. oceanica* NCP, NAP and NAP when standardised by canopy height. Tukey post hoc tests were used to test the differences between the seasons.

2.5.2 Influence of environmental conditions on *P. oceanica* NAP

The mean PME miniDOT water temperature was calculated across each incubation period. HOBO logger irradiance (lux) were converted to PAR (photosynthetic active radiation), according to the conversion factor appropriate to the light source ‘daylight’ (Thimijan and Heins, 1983). The Instantaneous PAR with the unit $\mu\text{mol s}^{-1} \text{m}^{-2}$ is used to calculate Daily Light integral (DLI) given the time interval in seconds (t) between each PAR reading over each 24-hour period:

$$\text{DLI} = \Sigma \Delta t \times \text{PAR}$$

The DLI is presented with the unit $\text{mol m}^{-2} \text{d}^{-1}$.

To examine the change in *P. oceanica* NAP relative to the changing light environment, the photosynthesis-irradiance relationship was fit with a hyperbolic tangent function (Jassby and Platt, 1976), modified to account for respiration (Rheuban et al., 2014), as used for the seagrass species in question (Koopmans et al., 2020). The model fitted was:

$$\text{Flux} = P_{\max} \tanh \frac{I}{I_k} - R_l$$

where P_{\max} is the maximum photosynthetic rate, I_k is the saturation irradiance, and R_l is respiration. The parameters were estimated by non-linear least squares approach (nls function, R version 3.5.1), estimating approximate start values. The irradiance compensation point is the irradiance at which net oxygen production equals zero.

As there was no significant interaction between temperature and seasonal NAP ($F_{1,4} = 0.189$, $P < 0.05$), or significant effect when temperature is assessed as a covariate for change in seasonal NAP by ANCOVA ($F_{1,6} = 2.1154$, $P < 0.05$) (ANNEX 3). The maximal model to assess for significant effect of temperature on NAP was by ANOVA.

2.5.3 Annual *P. oceanica* productivity

Annual NAP and NCP was estimated by scaling up the three average seasonal daily NAP and NCP measurements; the November measurement was chosen to represent the period between October to January (123 days), April represented the period from February to May (120 days) and July represented June to September (122 days):

$$\text{NAP}_{\text{Annual}} = (\text{NAP}_{\text{Daily(Nov)}} \times 123) + (\text{NAP}_{\text{Daily(Apr)}} \times 120) + (\text{NAP}_{\text{Daily(Jul)}} \times 122)$$

$$\text{NCP}_{\text{Annual}} = (\text{NCP}_{\text{Daily(Nov)}} \times 123) + (\text{NCP}_{\text{Daily(Apr)}} \times 120) + (\text{NCP}_{\text{Daily(Jul)}} \times 122)$$

2.5.4 Influence of spatial dynamics on *P. oceanica* NAP

A Welch’s t-test, to account for unequal sample size, is applied to determine if the NAP differs between the central meadow area and the meadow edge. Then linear regressions are applied to the summer NAP measurements to determine if the difference in central meadow and meadow edge relate to changes in plant surface area, percentage cover, shoot density and blade length. Comparison

of the R^2 values provides an understanding of which seagrass biometric account for the variation in NAP and are the best spatial predictor for *P. oceanica* NAP.

2.5.5 Species specific productivity comparison

The comparative species-specific NAP data conforms to normal distribution ($W_{1,7} = 0.922$, $P > 0.05$) and an ANOVA applied to assess if there was a significant difference in NAP between seagrass species during summer. The NAP standardised for plant surface area also confirmed to normal distribution ($W_{1,7} = 0.968$, $P > 0.05$). An ANOVA was applied, and Tukey post hoc tests confirmed any pairwise differences between species.

3 Results

3.1 Seasonal *P. oceanica* NCP and NAP

The *P. oceanica* meadow NCP is greater than the unvegetated control in every season (Fig. 3A). In autumn, the *P. oceanica* community is overall heterotrophic, as CR is greater than GPP (GPP:CR ratio < 1 , Fig. 3B). However, the oxygen deficit is greater in the community without seagrass (NCP $\bar{x} = -3.4$, $SD \pm 8.38$ O₂ mmol m⁻² d⁻¹), than in the *P. oceanica* community (NCP $\bar{x} = -2.0$, $SD \pm 3.24$ O₂ mmol m⁻² d⁻¹) (Fig. 3A). There is a significant seasonal influence on *P. oceanica* NCP ($F_{2,7} = 10.924$, $P < 0.05$). Pairwise post hoc comparisons show NCP is significantly higher in spring (NCP $\bar{x} = 8.3$, $SD \pm 3.12$ O₂ mmol m⁻² d⁻¹) and summer (NCP $\bar{x} = 7.8$, $SD \pm 2.03$ O₂ mmol m⁻² d⁻¹), compared to autumn when it is heterotrophic (Tukey: $P < 0.05$). There is no significant difference in NCP between the spring and summer.

During the transition from autumn to spring there is a large increase in GPP (GPP $\bar{x} = 20.2$, $SD \pm 7.81$ O₂ mmol m⁻² d⁻¹ to GPP $\bar{x} = 68.5$, $SD \pm 47.85$ O₂ mmol m⁻² d⁻¹, Fig. 3C) and a simultaneous increase in CR from autumn into spring (CR $\bar{x} = 22.2$, $SD \pm 10.12$ O₂ mmol m⁻² d⁻¹ to CR $\bar{x} = 60.2$, $SD \pm 44.80$ O₂ mmol m⁻² d⁻¹, Fig. 3D). However, the increase in GPP is greater than the increase in CR transitioning the *P. oceanica* meadow from heterotrophic in autumn to autotrophic in spring (GPP:CR ratio > 1 , Fig. 3B). The *P. oceanica* meadow stays in an autotrophic state in the Summer (GPP:CR ratio > 1 , Fig. 3B) registering its highest GPP (GPP $\bar{x} = 94.1$, $SD \pm 74.22$ O₂ mmol m⁻² d⁻¹, Fig. 3C), but also its highest CR (CR $\bar{x} = 86.4$, $SD \pm 74.21$ O₂ mmol m⁻² d⁻¹, Fig. 3C). The increase in GPP is not as great as the increase in CR, therefore the GPP to CR ratio and NCP is lower in summer compared to spring (Fig. 3B). Further to this the organisms present within the unvegetated control chambers have a higher metabolic rate in the summer (NCP $\bar{x} = -3.2$, $SD \pm 0.81$ O₂ mmol m⁻² d⁻¹) compared to the spring (NCP $\bar{x} = -0.4$, $SD \pm 0.71$ O₂ mmol m⁻² d⁻¹, Fig. 3A).

Whilst the *P. oceanica* NCP is highest in spring, *P. oceanica* NAP is highest in summer (NAP $\bar{x} = 10.9$, $SD \pm 3.03$ O₂ mmol m⁻² d⁻¹) (Fig. 4A). There remains a significant seasonal difference in *P. oceanica* NAP ($F_{2,7} = 8.3885$, $P < 0.05$). However, Tukey post hoc comparisons found NAP is only significantly lower in autumn compared to summer (Tukey: $P < 0.05$). Although there is an observational difference in NAP between the autumn (NAP $\bar{x} = 1.4$, $SD \pm 3.24$ O₂ mmol m⁻² d⁻¹) and spring (NAP $\bar{x} = 8.7$, $SD \pm 3.12$ O₂ mmol m⁻² d⁻¹) the difference is not significant. There remains no significant difference in NAP between the periods of highest productivity in spring and summer.

The canopy height within the edge *P. oceanica* habitat is lowest during autumn ($\bar{x} = 15.4$, $SD \pm 2.84$ cm), this increases into spring ($\bar{x} = 22.3$, $SD \pm 4.77$ cm) and peaks during summer ($\bar{x} = 25.1$, $SD \pm 5.90$ cm). When the *P. oceanica* NAP is standardised by the seagrass canopy height in the corresponding season, autumn still maintains the lowest NAP, but it highlights that *P. oceanica* NAP

is more variable within autumn (NAP \bar{x} = 0.07, SD \pm 0.221 O₂ mmol m⁻² cm⁻¹ d⁻¹) (Fig. 4B). In comparison *P. oceanica* NAP in Summer (NAP \bar{x} = 0.43, SD \pm 0.066 O₂ mmol m⁻² cm⁻¹ d⁻¹), is less varied and similar to spring (NAP \bar{x} = 0.40, SD \pm 0.125 O₂ mmol m⁻² cm⁻¹ d⁻¹) (Fig. 4B). Thus, the significant seasonal difference in NAP remains, between autumn and summer ($F_{2,7}$ = 6.3149 P < 0.05). There is no significant difference in NAP between autumn and spring or spring and summer.

3.2 Influence of environmental conditions on *P. oceanica* NAP

The daily light integral ranged from 4.7 mol photons m⁻² d⁻¹ during autumn to 25.2 mol photons m⁻² d⁻¹ in the summer. Variation in environmental conditions within each season allowed for some overlap in the irradiance encountered between seasons and the *P. oceanica* NAP resembled a saturation curve when plotted as a function of irradiance (Fig. 5A), as did the curve for NAP standardised by canopy height (Fig. 5B). The DLI conditions in autumn (Min = 4.7 mol photons m⁻² d⁻¹) sit near the predicted irradiance compensation point (IC = 4.6 mol photons m⁻² d⁻¹) as such within the autumnal DLI range there are conditions where the plant may only just or may not maintain metabolic requirements due to low irradiance. In spring the seagrass transitions between light limiting and light saturating levels. The light saturating conditions occur throughout summer, when NAP (NAP \bar{x} = 10.9, SD \pm 3.03 O₂ mmol m⁻² d⁻¹ Fig. 4A) sits near the predicted maximum (10.12 O₂ mmol m⁻² d⁻¹, Fig. 5A). The period of summer light saturation corresponds to when NAP is less varied whilst changes in canopy height across the year are also accounted for (Fig. 4B). The changes in the DLI account for 71% of the variation in *P. oceanica* NAP.

The water temperature was lowest in spring (\bar{x} = 17.7, SD \pm 0.13 °C), increasing to a summer high (\bar{x} = 23.6, SD \pm 0.40 °C). The autumn temperatures encountered sat at a relative mid-point (\bar{x} = 20.7, SD \pm 0.45 °C) between spring and summer, yet the NAP was lowest in autumn. Therefore, the change in NAP was not relative to the change in water temperature and not a significant factor as main effects (ANNEX 3), with season ($F_{1,6}$ = 2.1154, P > 0.05) or without season (Temperature $F_{1,8}$ = 0.7265, P > 0.05).

3.3 Annual productivity of *P. oceanica* edge habitat

The annual NAP of the *P. oceanica* edge habitat in this meadow was estimated at 2.6 C mol m⁻² yr⁻¹. However, the organisms present within the *P. oceanica* meadow have their own metabolic requirements (NCP without seagrass present -0.85 C mol m⁻² yr⁻¹), as such the net carbon gain at the community level for this *P. oceanica* meadow was less than the NAP produced annually by the *P. oceanica* (NCP = 1.7 C mol m⁻² yr⁻¹).

3.4 Influence of spatial dynamics on *P. oceanica* NAP

The *P. oceanica* NAP in summer is significantly different between the central areas of the meadow and the meadow edge ($t_{(1,4)}$ = 3.7647, P < 0.05). (Fig. 6). The *P. oceanica* NAP is highest in the central areas of the meadow (NAP \bar{x} = 21.7, SD \pm 4.21 O₂ mmol m⁻² d⁻¹) and approximately two-fold that of the edge habitat (NAP \bar{x} = 10.9, SD \pm 3.03 O₂ mmol m⁻² d⁻¹). The *P. oceanica* NAP increases relative to the plant surface area of *P. oceanica* (LM: $F_{1,5}$ = 132.5, P < 0.01) (Fig. 6A). The plant surface area was the best predictor of NAP and accounted for more than 96 % of the variation in NAP. As standalone biometrics of seagrass meadow metabolism, shoot density (m²), blade length (cm) and seagrass cover (%), were all significant predictors of NAP (shoot density, LM: $F_{1,5}$ = 21.13, P < 0.01; blade length, LM: $F_{1,5}$ = 12.98, P < 0.05; percentage cover, LM: $F_{1,5}$ = 8.78, P < 0.05) but they only accounted for 81 %, 72 % and 64 % (Fig. 6B, C and D) of the variation in NAP of the *P. oceanica*.

3.5 Species specific productivity

The seagrass benthic communities at 7m depth are all heterotrophic at the community level, ranging from the least heterotrophic with *H. stipulacea* present (NCP \bar{x} = -4.8, SD \pm 4.7 O₂ mmol m⁻² d⁻¹), followed by the community with *P. oceanica* present (NCP \bar{x} = -7.5, SD \pm 8.9 O₂ mmol m⁻² d⁻¹) and finally the most heterotrophic the *C. nodosa* dominant community (NCP \bar{x} = -10.5, SD \pm 6.0 O₂ mmol m⁻² d⁻¹) (Fig. 7A). The *H. stipulacea* NAP appears to be the highest comparatively and the only seagrass species to on average maintain positive carbon balance (O₂ > 0). However, variation in NAP for *H. stipulacea* (NAP \bar{x} = 2.1, SD \pm 4.7 O₂ mmol m⁻² d⁻¹) and *P. oceanica* (NAP \bar{x} = -3.6, SD \pm 8.9 O₂ mmol m⁻² d⁻¹) show both species fluctuate between positive and negative carbon balance within this area of the bay in Summer (Fig. 7B). Whilst *C. nodosa* NAP is the lowest and consistently negative (NAP \bar{x} = -8.6, SD \pm 6.0 O₂ mmol m⁻² d⁻¹), however, there was no significant statistical difference between species NAP (7B) ($F_{2,14}$ = 3.3806, P > 0.05).

The difference spatially in above ground vegetative material between species is demonstrated through the increasing total plant surface between species from *H. stipulacea* (\bar{x} = 5.1, SD \pm 1.4 x10⁻³ cm² m⁻²), to *C. nodosa* (\bar{x} = 10.6, SD \pm 9.4 x10⁻³ cm² m⁻²) and culminating in *P. oceanica* (\bar{x} = 17.6, SD \pm 7.3 x10⁻³ cm² m⁻²). However, the plant surface area is also more uniform for *H. stipulacea* spatially across the habitat sampled, in comparison the plant surface area of both *C. nodosa* and *P. oceanica* show higher variation between samples (Fig. 7C). Standardising the seagrass NAP for the relative plant surface area within each sample, alters the relative differences between species NAP (7B and D). The NAP standardised for plant surface area highlights a significant difference does occur between species ($F_{2,14}$ = 4.1922, P < 0.05). Tukey post hoc comparisons show *C. nodosa* NAP when standardised by plant surface area (NAP \bar{x} = -0.0012, SD \pm 0.0007 O₂ mmol cm⁻² d⁻¹) to be significantly lower than *H. stipulacea* (NAP \bar{x} = 0.0004, SD \pm 0.0011 O₂ mmol cm⁻² d⁻¹) (Tukey: P < 0.05). But there was no significant difference between the native species *C. nodosa* and *P. oceanica* NAP (NAP \bar{x} = -0.0004, SD \pm 0.0009 O₂ mmol cm⁻² d⁻¹). There was also no significant difference between *H. stipulacea* and *P. oceanica* NAP.

4 Discussion

4.1 Seasonal *P. oceanica* NCP and NAP

We found strong seasonality in *P. oceanica* productivity, at both the community (NCP Spring \bar{x} = 8.3, SD \pm 3.12; Autumn \bar{x} = -2.0, SD \pm 3.24 O₂ mmol m⁻² d⁻¹) and plant level (NAP Summer \bar{x} = 10.9, SD \pm 3.03; Autumn \bar{x} = 1.4, SD \pm 3.24 O₂ mmol m⁻² d⁻¹), which reinforces the need for seagrass carbon sequestration studies to consider year-round dynamics, thereby ensuring carbon storage estimates are not exaggerated by only reflecting summer peaks. Similar conclusions from studies of net ecosystem productivity in freshwater lakes (Brentrup et al., 2020), and temperate saltmarshes (Vazques Lule & Vargas 2021), suggest that this should now be the norm.

Multi-season approaches enable the assessment of seasonal drivers such as irradiance (Gazeau et al., 2005; Champenois and Borges, 2012). During autumn, the overall irradiance reaching the plant is lower as the daylight period is shorter, the likelihood of cloud cover increases, and increased frequency of stormy conditions results in greater hydrodynamic activity reducing in-water visibility. Subsequently autumnal photosynthetic activity occurs when conditions are light limiting, promoting a greater variability in productivity. In contrast light saturating levels present throughout summer, lead to a lower *P. oceanica* photosynthetic efficiency. Long-term exposure to saturating irradiance decreases plant productivity and can lead to photoinhibition (Ralph and Burchett, 1995), though a decline from photoinhibition is not considered in this study. The consecutive days of light saturating

irradiance which occur in summer represent an increasing likelihood that photoinhibition would occur. However, shallow water *P. oceanica* have been demonstrated to adapt to high light conditions, by specific photoacclimative and photoprotective responses; significant enrichment of up-regulated transcripts associated with light-dependent reaction of photosynthesis (e.g., light-harvesting proteins); and modifications in the structure of photosystems and tocopherol biosynthesis activation in the summer (Dattolo et al., 2014). This is potentially why a strong negative photoinhibition response was not observed in our seasonal metabolism measurements. The saturation response alone has important considerations for modelling carbon sequestration capacity, re-emphasising the importance of seasonality dependent on the concurrent irradiance levels. Physical disturbance to seagrass meadows such as autumnal and winter storms will seasonally reduce the available photosynthetically active plant material, i.e., as storms tear seagrass blades. Although we have considered seasonal changes in seagrass metabolism and productivity, we have not directly measured biomass export over winter, which can be vast enough seagrass exported onshore forms large banquettes (Gomez-Pujol et al., 2013). This needs to be considered in future estimates.

Given *P. oceanica* meadows within the eastern region of the Aegean Sea have been noted to be fragmented in response to elevated sea surface temperatures (Chefaoui et al., 2017), it is surprising we found no statistically significant influence of temperature on the productivity of shallow water *P. oceanica*. Other temperate seagrass species have been shown to transition to a negative carbon balance when exposed to high Summer water temperatures (Marsh Jr. et al., 1986). Although our highest average daily water temperature was above optimal thermal conditions (23.9 vs 17 - 20 °C) for *P. oceanica* (Champenois and Borges, 2018), this was well below *P. oceanica* critical temperature limits (> 29 °C). The *in-situ* nature of this study does not allow for the independent control of water temperature or light intensity encountered at any time, and so they are often correlated making it challenging to differentiate between them. Shallow water *P. oceanica* has a greater tolerance to increased water temperatures than deeper *P. oceanica* (Marin-Guirao et al., 2016). This heat acclimation in shallow *P. oceanica* emerges by stabilising respiration, thereby establishing respiratory homeostasis (i.e., balanced photosynthetic carbon gains and respiratory carbon losses; Marin-Guirao et al., 2016). This suggests that shallow *P. oceanica* may maintain net productivity gains at higher water temperatures for longer periods. It is also possible that the *P. oceanica* within the Eastern Mediterranean may have a greater acclimation to increased water temperature, as the Eastern Mediterranean basin has been exposed to higher sea surface temperatures for longer than the Western basin (since 1985) (Nykjaer, 2009). Ultimately any temperature effect on *P. oceanica* metabolism in this study was confounded by the strong irradiance effect and thus could not be disentangled.

4.2 Annual *P. oceanica* NCP and NAP

The annual net community productivity for *P. oceanica* meadows in our study was lower than recorded for two *P. oceanica* meadows previously studied in the western region of the Aegean Sea, even one negatively impacted by eutrophication (4.83 C mol m⁻² yr⁻¹) (Apostolaki et al., 2010), however this study used smaller chambers, which are more likely to cause oxygen saturation which can overestimate productivity. The annual NCP in this study was conservative as it was obtained from patchy *P. oceanica* edge habitat, this underestimates the meadows overall carbon sequestration ability which consisted of both dense and patchy habitat. However, in shallow water meadows, patchy edge habitat is a more prominent feature (Montefalcone et al., 2009), therefore it is an appropriate conservative annual NCP estimate. Although we found meadow centre *P. oceanica* productivity was two-fold that of the edge habitat, the overall meadow autochthonous carbon sequestration potential lies between 1.7 - 3.4 C mol m⁻² yr⁻² which remains lower than other studies

have found. Despite a low annual NCP estimate our shallow water *P. oceanica* remained autotrophic actively sequestering carbon, which if transitioned to an unvegetated state would result in it becoming overall heterotrophic, a carbon source.

4.3 Spatial dynamics of *P. oceanica* NAP

Metabolism measurements are typically presented as an area-based unit (Apostolaki, et al., 2010; Duarte et al., 2010; Olive et al., 2015). We have shown Net Apparent Productivity to be affected by spatial variation in plant aboveground biomass, this would suggest that reported estimates may benefit from additional information on canopy height and shoot density. Estimates should potentially move away from denoting discrete areal measurements for meadows. Instead acquiring and mapping large scale data on canopy height (shoot size) and shoot density to enable comprehensive habitat wide autochthonous carbon sequestration estimates. Acquiring seagrass biometric data to support productivity estimates requires greater effort than collating presence-absence or coverage data, but simple plant biometrics of shoot density and leaf area index have been demonstrated as standalone predictors of carbon sequestration and community composition (Samper-Villarreal et al., 2016; Collier et al., 2021). High shoot density in shallow areas with a high degree of patchiness creates very specific nest-like patterns, these have their highest shoot density in the centre and decrease in density radially towards the edge of the patches (Zupo et al., 2005). Therefore, this distinction in spatial productivity between edge and central areas of *P. oceanica* may be a specific characteristic to consider for shallow meadows because overall shoot densities are highest in the shallow and decrease with depth (Olesen et al., 2002; Zupo et al., 2005). More work would be needed to determine if the same distinctions in shoot density and subsequent primary productivity exist between the edge and central areas of *P. oceanica* meadows in deeper waters. The use of chambers with a larger benthic surface area has a greater representation of community-scale metabolism, as trialled for heterogenic coral reef habitats (Yates and Halley, 2003). The use of a larger benthic chamber for *P. oceanica* metabolism measurements in this study (1m diameter) comprises community metabolism measurements influenced by fine scale variation in shoot density, compared to previous studies using smaller chambers (0.18m diameter), which house only a few shoots at similar densities within the benthic chamber (Gazeau et al., 2005; Barron et al., 2006).

The carbon sequestration potential of seagrass meadows ultimately relates to how much of the organic material produced by the seagrass, i.e. the productivity, accumulates as refractory material in the seagrass meadow. Although the patchiness of shallow meadows may render them less effective at attenuating currents and promoting settlement of organic and inorganic particles, they still actively contribute to carbon acquisition within the carbon cycle. Given a total of 7139.92 km² of *P. oceanica* is estimated to occur in the Eastern Mediterranean Basin (Telesca et al., 2015), and seagrass coverage within Greek waters has been estimated at 2619 km² (Topouzelis et al., 2018), *P. oceanica* within Greek waters represents around 36.7% of the total *P. oceanica* in the Eastern Mediterranean. The quantity of *P. oceanica* habitat within the Aegean Sea and Eastern Mediterranean means it represents a substantial blue carbon habitat irrespective of its fragmented nature (Chefoui et al., 2018). The further loss of this habitat would risk climate current change targets therefore seagrass meadows need distinct conservation management strategies given their proximity to current threats including impacts from human infrastructure and coastal activities (Giakoumi et al., 2015).

4.4 Species specific productivity

We found *P. oceanica* meadow to be overall annually autotrophic, however *P. oceanica* at 7m depth fluctuated between positive and negative carbon balance. Such heterogeneity over a small scale within the same bay highlights that adjacent *P. oceanica* patches may simultaneously undergo growth

and loss. Given *P. oceanica* at 7m depth was particularly short (Plant surface area $\bar{x} = 17.6$, $SD \pm 7.3 \times 10^{-3} \text{ cm}^2 \text{ m}^{-2}$) relative to the other depths where the plant surface area was higher (Ranging from $37.6 - 293.0 \times 10^{-3} \text{ cm}^2 \text{ m}^{-2}$) and given plant surface area acts as a predictor of productivity, lower productivity is expected. It must also be considered that our results for this comparison are influenced by differences in chamber area, volume, and incubation time.

Based on our productivity assessments alone, the non-native *H. stipulacea* was not competitive over *P. oceanica*. This is akin to other studies which suggest a limited capacity to competitively displace *P. oceanica* (Williams, 2007; Winters et al., 2020), although *H. stipulacea* can opportunistically colonise space previously occupied by *P. oceanica*. With regression of *P. oceanica* meadows well documented (Telesca et al., 2015), alongside *H. stipulacea* range expansion (Georgiou et al., 2016), maintenance of Mediterranean seagrass meadow areal cover through replacement of current seagrass species cover is likely. *H. stipulacea* productivity in this introduced range of the Mediterranean is comparatively lower than its native range, given irradiance was the driving factor in *H. stipulacea* productivity within its home range in the Gulf of Aquaba (Cardini et al. 2018). We found *H. stipulacea* productivity to be comparable to patchy short *P. oceanica* (NAP $\bar{x} = 2.1$, $SD \pm 4.7$ vs -3.6 , $SD \pm 8.9 \text{ C mmol m}^{-2} \text{ d}^{-1}$). The carbon sequestration capacity of *H. stipulacea* has been estimated to be greater than *P. oceanica* in the Eastern Mediterranean (Wesselmann et al., 2021), but not greater than *P. oceanica* carbon sequestration estimates in the Western Mediterranean (Mazarrasa et al., 2017). Given its tropical origins, under future climate change predictions of a warming Mediterranean *H. stipulacea* has potential to increase its productivity, whilst the same cannot be said for *P. oceanica*.

Whilst *H. stipulacea* has persisted within the Mediterranean since 1984 and expanded its known range (Fritsch, 1895; van der Velde and den Hartog, 1992; Gambi et al., 2009; Tsiamis et al., 2010; Sghaier et al., 2011; Georgiou et al., 2016). *H. stipulacea* is known to disappear and recolonise as per a metapopulation (Gambi et al., 2018), making the persistence of its sedimentary carbon stocks questionable. While questions remain, it would appear the non-native *H. stipulacea* is already making positive contributions to community blue carbon in the eastern Aegean.

The replacement of *P. oceanica* by *C. nodosa* is well documented (Montefalcone et al., 2006; Montefalcone et al., 2007). This replacement is typically considered a result of *C. nodosa* being more tolerant to varying environmental conditions. Compared against six dominant native Mediterranean macrophytes *C. nodosa* presented the highest thermal optima (Savva et al., 2018). But the substitution of *P. oceanica* for *C. nodosa* is not exclusive, replacement of *P. oceanica* dead matte to algae dominated *Caulerpa spp.* habitats also occurs (Montefalcone et al., 2007). *C. nodosa* may not always be the ‘winner’ following loss or fragmentation of *P. oceanica* meadows. We found when the biomass of *H. stipulacea* and *C. nodosa* are comparable, *H. stipulacea* can hold a metabolic advantage over *C. nodosa*. In the Southern Mediterranean replacement of *C. nodosa* by *H. stipulacea* under warmer environmental conditions can occur (Sghaier et al., 2014). Very shallow water will be readily heated by the sun, in the North Mediterranean this may provide warmer conditions closer to the tropical native range of *H. stipulacea*. Within our study site, but at a shallower location (0.5m depth), *H. stipulacea* was found mixed within dense *C. nodosa* beds and growing multiple paired leaves off a vertical stem (Pers. obv., ANNEX 4). This is important given lateral growth is more typical of this species (Posluszn and Tomlinson, 1990; Winters et al., 2020). Vertical growth would give *H. stipulacea* the potential to increase its relative biomass and gain a competitive advantage over *C. nodosa*. *C. nodosa* cover is predicted to reduce under future warming scenarios due to environmental changes (circa -46.5%) (Chefaoui et al., 2018). As per the interactions with

622 *P. oceanica*, a warming climate will lead to a faster and larger decline in *C. nodosa*, beyond the
623 current predictions that do not yet account for competition with *H. stipulacea*.

624 **4.5 Conclusions**

625 If natural ecosystems such as seagrass meadows are to play a part in mitigating increases in
626 anthropogenic CO₂ emissions spatio-temporal variability in primary productivity of seagrass
627 meadows needs to be accounted for. This study demonstrates that not all seagrass habitat is equal in
628 its potential to sequester autochthonous carbon, but the variability can in part be accounted for;
629 seasonally and spatially, using environmental predictors such as irradiance and plant biometrics such
630 as plant surface area. Alongside this the non-native *H. stipulacea* contributes to community Blue-
631 Carbon and has a metabolic advantage over *C. nodosa* under current conditions when both species
632 biomass are comparable. Under a warming Mediterranean this may promote direct competition
633 between the two species. Mediterranean seagrass meadow species composition, the condition of these
634 meadows and the potential interactions between seagrass species must be better understood for
635 accurate predictions of future seagrass community Blue Carbon productivity.

636 **5 Conflict of Interest**

637 The authors declare that the research was conducted in the absence of any commercial or financial
638 relationships that could be perceived as a potential conflict of interest.

639 **6 Author Contributions**

640 EW, TC, TW, CA, TT and AM contributed to the study conception and design. EW and CA led the
641 field-based studies, supported by TT and AM. TW led a preliminary pilot study integral to our final
642 experimental design. EW and TC led on data analysis and the manuscript first draft. All authors were
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654 **9 Supplementary Material**

655 The Supplementary Material are uploaded separately for submission.

656 **10 Data Availability Statement**

657 Datasets are available from the corresponding authors and will become available on the University of
658 Essex data repository.

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887 12 Figure captions

888 **Figure 1.** Large benthic chamber setup (A) Deployed in autumn over *P. oceanica* edge habitat (B)
889 Deployed in summer over dense central *P. oceanica* meadow. (C) Setup of the internal water pump
890 (foreground) within a benthic chamber over control unvegetated habitat. External submersible pump
891 battery pack in background. (D) Illustration of large benthic chamber set-up. Photographs © Emma A
892 Ward, Illustration © Tom Wade.

893 **Figure 2.** Small benthic chamber setup (A) Deployed over short patchy *P. oceanica* habitat (B)
894 Aerial view above patchy *P. oceanica* chambers deployed. Photographs; A and B © Jente van
895 Langerak.

896 **Figure 3.** (A) Net community productivity (NCP), (B) GPP:CR Ratio, (C) Gross primary
897 productivity (GPP) and (D) Community respiration (CR) for the *P. oceanica* meadow edge (grey)
898 and control chamber (White), autumn (November), spring (April) and summer (July). Error bars
899 represent standard deviation (autumn and spring n = 3, summer n = 4).

900 **Figure 4.** (A) The *P. oceanica* Net Apparent Productivity (NAP) given the season, (B) The
901 *P. oceanica* NAP when standardised by seagrass canopy height in the given season. Autumn and
902 spring n = 3, summer n = 4. Error bars represent standard deviation.

903 **Figure 5** (A) *P. oceanica* NAP ($R^2 = 0.713$) and (B) *P. oceanica* NAP standardised by canopy length
904 ($R^2 = 0.711$), as a function of PAR given as the Daily Light Integral. The irradiance compensation
905 point (I_c) was 4.6 mol photons $m^{-2} d^{-1}$. Replicate season denoted for visual reference autumn (●),
906 spring (●) and summer (●).

907 **Figure 6.** Net apparent productivity (NAP) of the *P. oceanica* in the central meadow (grey) and
908 meadow edge (Black) in relation to (A) *P. oceanica* blade surface area (B) *P. oceanica* shoot density
909 (m^{-2}) (C) *P. oceanica* blade length (cm) and (D) *P. oceanica* cover (%). Central meadow n = 3,
910 meadow edge n = 4. Significant linear regression from edge and central meadow combined * = $P <$
911 0.05, ** = $P < 0.01$. Grey bands reflect the 95% confidence level interval for predictions from a
912 linear model.

913 **Figure 7.** (A) Net community productivity (NCP), (B) Net apparent productivity (NAP), (C) Leaf
914 surface Area (D) NAP standardised by sampled plant surface area, for the three seagrass species
915 *H. stipulacea* (n = 6), *C. nodosa* (n = 5) and *P. oceanica* (n = 6). Error bars represent standard
916 deviation.